

Substrate specificity of the MBP-SelU fusion protein (Escherichia coli tRNA 2-selenouridine synthase)

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Transfer RNAs (tRNAs) constitute a unique group of RNA oligonucleotides, which is characterized by the highest content of modified nucleosides. The 5-substituted nucleosides, like 2-thiouridines (R5S2U), S-geranyl-2-thiouridines (R5geS2U), and 2-selenouridines (R5Se2U), found in bacterial tRNAs specific for Lys, Glu, and Gln at the first position of their anticodons (a position 34, the wobble position), play a pivotal role in the decoding of genetic information. Bacterial tRNA 2-selenouridine synthase (SelU, MnmH) exerts a two-step post-transcriptional transformation of (c)mnm5S2U-tRNA to (c)mnm5Se2U-tRNA via a (c)mnm5geS2U-tRNA intermediate.¹ In the current studies we used an MBP-SelU fusion protein, with a maltose binding protein (MBP) tag attached to the N-terminus of the wild type SelU synthase. MBP-SelU (84.4 kDa) shows high activity in the S-geranylation (> 90%) and selenation (~100%) of model 17-mer oligonucleotides mimicking the anticodon-stem-loop (ASL) of tRNA^{Lys}. These models contained at the position corresponding to the wobble position in tRNA anticodon either an S2U or geS2U unit, respectively. The MBP-SelU activity and the minimum length of the RNA oligonucleotide substrate, as well as the influence of the position of modified nucleoside and flanking sequences on the enzyme/substrate recognition were determined. We confirmed that the MBP-SelU protein binds the bacterial tRNA^{Lys}, tRNA^{Glu} and tRNA^{Gln}. Using Microscale Thermophoresis (MST) technique the constants for binding of Cy3-labeled 17-mer oligonucleotides (containing U, S2U, geS2U or Se2U) to MBP-SelU protein were determined. The geS2U-RNA exhibits the highest affinity for MBP-SelU ($K_d=3.946 \pm 0.41 \mu\text{M}$) in comparison with S2U-RNA ($K_d=18.54 \pm 3.01 \mu\text{M}$), Se2-RNA ($K_d=27.33 \pm 4.33 \mu\text{M}$).

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¹ Sierant M et al. (2018) FEBS Lett. 592, 2248-2258